

MOLECULAR CONFORMATION OF ERABUTOXIN b; ATOMIC COORDINATES AT 2.5 Å RESOLUTION

M. R. Kimball, A. Sato*, J. S. Richardson**, L. S. Rosen, and B. W. Low†
Department of Biochemistry, Columbia University, 630 W. 168 St., N.Y., N.Y. 10032

Received April 16, 1979

SUMMARY: Atomic coordinates determined from a 2.5 Å electron density map are given for erabutoxin b, a sea snake venom postsynaptic neurotoxin. The principal structural features, anti-parallel β pleated sheet, β bulge and β bends are described. The erabutoxin b structure is discussed as structural prototype of this class of homologous curare-mimetic neurotoxins from both land and sea snakes.

In this communication we provide atomic coordinates for erabutoxin b, a sea snake venom neurotoxin, based on a 2.5 Å electron density map and describe major features of the molecular conformation and stereochemistry of this single chain protein (M_r 6861) considered as prototype for a whole class of neurotoxins. Erabutoxin b, from the sea snake Laticauda semifasciata, is one of nearly 50 homologous neurotoxins from both land (elapid) and sea (hydrophid) snake venoms which bind to the cholinergic receptor at the postsynaptic membrane and block neuromuscular transmission in a nondepolarizing curare-mimetic mode.

The main structural features of this short chain toxin observed at a lower resolution were described earlier, and a preliminary discussion of the distribution of some common invariant and conservatively-substituted[†] residues given (1). The significance of the erabutoxin b structure as prototype was recognized and a reactive site region described (1,2). There are two toxin series, short with 60-62 and long with 71-74 residues, with 18 invariant and 7 conservatively-substituted positions in common, which include four invariant disulfide linkages. These latter play a unique and evident struc-

*Present address: Department of Chemistry, Tohoku University, Sendai, Japan.

**Permanent address: Department of Anatomy, Duke University, Durham, N.C.

†To whom all correspondence should be addressed.

†Conservative substitution is used here to define conservation of residue type, i.e., Ser/Thr-hydroxyl group, Glu/Asp-carboxyl group, Val/Leu/Arg-hydrophobic group and Tyr/His/Trp-aromatic group.

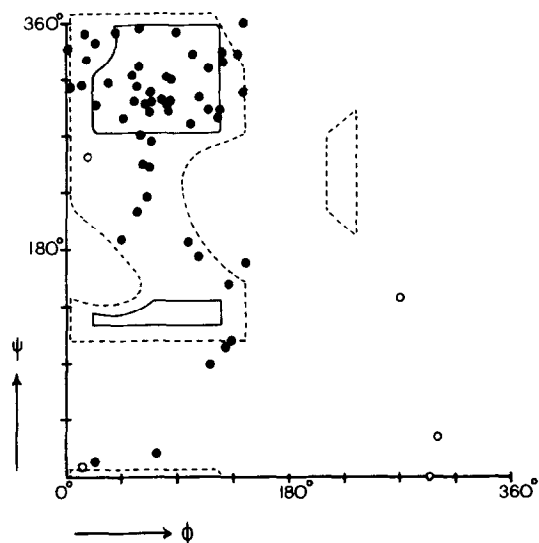


Fig. 1. Ramachandran angle plot for erabutoxin b showing "fully allowed" (—) and "outer limit" (---) boundaries. [○] Gly; [●] Other residues.

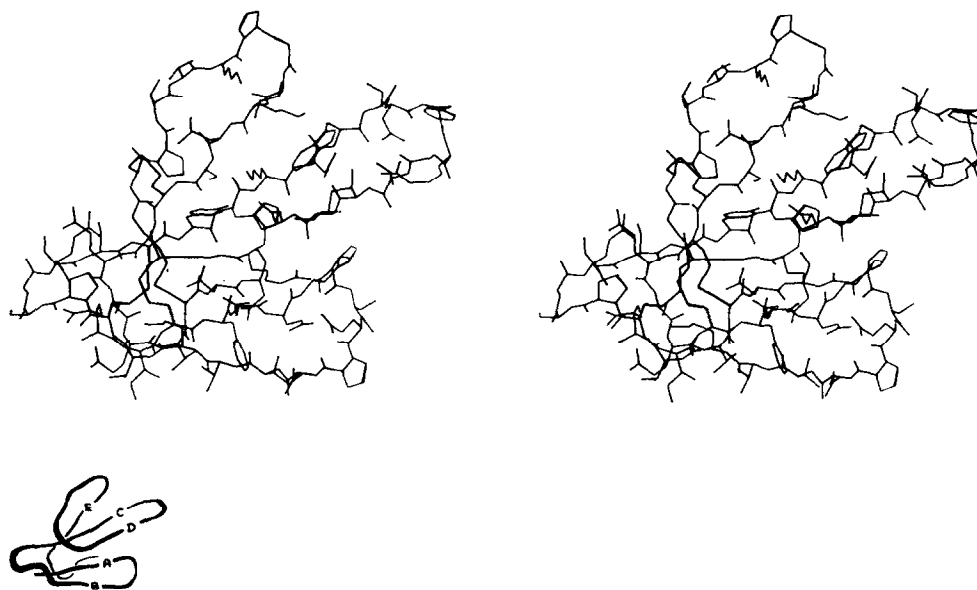


Fig. 2. Stereo drawing of erabutoxin b structure showing all the residues, produced by the GRIP-75 Molecular Graphics System.

tural role; both native conformation and toxicity are lost completely when they are reduced (3).

TABLE 1 Atomic Coordinates for Erabutoxin b.

10x 10y 10z				10x 10y 10z				10x 10y 10z				10x 10y 10z			
1 ARG				10 GLN				20 GLY				29 TRP			
N	114	74	93	N	277	71	242	N	53	109	102	N	268	206	241
CA	126	60	100	CA	270	61	250	CA	59	115	90	CA	268	218	250
CB	123	94	102	CB	257	66	255	C	68	127	94	CB	254	223	250
CG	134	103	97	CG	244	57	254	O	65	138	90	CG	243	214	258
CD	144	95	87	CD	230	62	257	21 GLU				CD1	242	214	271
NE	145	100	74	OE1	225	72	252	N	79	125	101	NE1	233	204	274
CE	135	99	65	NE2	223	55	266	CA	90	135	100	CE2	228	199	263
NEH2	136	104	53	C	264	51	239	CB	94	140	113	CD2	234	205	252
NEH1	124	92	67	O	263	53	227	CG	99	153	111	CE3	231	201	240
C	129	73	112	11 PRO				CD	104	150	124	CE3	221	191	238
O	122	73	122	N	264	38	243	OE1	107	171	124	C22	216	185	249
2 ILE				CD	265	32	256	OE2	105	151	134	C	277	216	262
N	141	66	111	CA	262	29	231	C	101	130	92	O	282	205	265
CA	140	61	123	CB	261	16	237	O	107	119	95	30 SER			
CB	154	47	120	CG	263	17	252	22 SER				N	279	228	267
CG2	157	39	133	C	250	34	224	N	104	139	83	CA	290	232	276
CG1	144	38	112	G	241	40	231	CA	114	137	72	CB	298	244	271
CD1	143	24	117	12 GLN				CB	112	149	62	OG	289	252	262
C	162	65	123	N	251	29	212	OG	107	160	69	C	285	236	290
O	171	60	116	CA	241	33	201	C	128	137	78	G	289	246	296
3 CYS				CB	240	28	188	O	138	135	70	31 ASP			
N	164	75	132	CG	261	34	185	23 SER				N	275	227	294
CA	177	81	134	CC	265	33	170	N	130	140	90	CA	270	229	307
CB	174	96	133	OE1	272	42	165	CA	143	145	94	CB	259	218	309
SG	170	101	115	NE2	262	22	163	CB	145	160	93	CG	259	205	315
C	183	75	147	C	228	27	203	OG	135	166	101	OD1	270	198	319
O	179	65	152	G	226	14	204	C	150	143	107	OD2	248	200	317
4 PHE				13 THR				O	145	135	116	C	282	222	314
N	193	82	152	N	218	35	204	24 CYS				O	292	218	307
CA	201	78	163	CA	203	31	205	N	161	149	111	32 PHE			
CB	216	79	160	CB	199	30	219	CA	168	144	123	N	285	225	328
CG	222	66	156	CG2	207	20	227	CB	172	130	118	CA	298	222	333
CD1	221	55	164	OG1	200	43	226	SG	159	119	117	CB	298	221	346
CE1	227	42	161	C	196	41	196	C	179	152	131	CG	285	218	350
CZ	234	41	149	O	201	51	191	O	183	162	127	CD1	272	220	346
CE2	235	52	140	14 THR				25 TYR				CE1	262	216	354
CD2	229	65	144	N	183	38	193	N	183	146	142	C2	264	209	365
C	198	86	176	CA	177	45	182	CA	188	154	153	CE2	277	207	370
O	202	98	177	CB	181	36	170	CB	174	161	159	CD2	287	212	362
5 ASN				CG2	173	23	168	CG	174	161	172	C	303	208	329
N	191	80	185	OG1	180	44	158	CD1	162	162	178	O	311	206	321
CA	188	85	198	C	161	45	183	CE1	162	162	192	33 ARG			
CB	177	76	205	O	155	36	190	CE	173	162	200	N	297	199	336
CG	169	84	215	15 LYS				OE1	173	162	213	CA	301	185	337
OD1	159	90	212	N	155	55	177	CE2	186	160	193	CB	288	176	335
ND2	173	83	227	CA	140	57	178	CD2	186	160	179	CG	282	169	346
C	200	84	208	CB	137	70	184	C	202	153	159	CE	267	162	343
O	200	87	220	CG	127	70	195	G	209	142	159	NE	260	163	330
6 GLN				CD	113	75	191	26 HIS				C2	249	171	328
N	212	79	203	CE	101	71	200	N	206	165	163	NEH2	243	173	317
CA	224	76	211	NZ	98	72	194	CA	219	168	169	NEH1	245	179	330
CB	235	70	202	C	132	57	165	CB	230	161	161	C	306	181	324
CG	247	73	207	G	138	62	155	CG	244	162	166	O	311	169	322
CD	254	71	199	16 THR				ND1	253	151	164	34 GLY			
OE1	240	68	190	N	121	51	163	CE1	264	155	170	N	305	189	314
NE2	267	72	198	CA	112	52	151	NE2	263	167	175	CA	312	187	301
C	229	89	218	CB	106	38	148	CD2	251	171	173	C	302	186	289
O	234	98	211	CG2	95	35	158	C	221	180	177	O	291	192	289
7 HIS				OG1	100	38	135	27 LYS				35 THR			
N	227	90	231	C	102	62	154	N	227	178	189	N	307	179	280
CA	234	100	238	O	97	64	165	CA	231	188	198	CA	301	177	266
CB	233	97	253	17 CYS				CB	219	195	205	CB	310	183	257
CG	239	107	261	N	99	71	145	CB	211	185	212	CG2	319	174	249
ND1	233	113	272	CA	92	84	149	CE	199	191	218	OG1	304	192	248
CE1	242	121	277	CB	101	96	146	CD	190	181	225	C	292	164	265
NE2	253	121	271	SG	120	95	145	NZ	178	188	231	O	297	153	260
CD2	252	112	261	C	78	87	144	C	243	184	207	36 ILE			
C	247	102	232	O	73	79	136	G	246	172	208	N	281	169	262
O	253	93	225	18 PRO				28 GLN				CA	269	162	259
8 SER				N	71	97	148	N	249	194	213	CB	257	168	267
N	257	107	232	CD	77	107	157	CA	261	195	221	CG2	259	164	268
CA	268	111	225	CA	58	100	143	CB	271	199	211	CG1	243	167	258
CB	271	120	232	CB	54	113	150	CG	263	189	211	CD1	240	153	256
OG	268	117	246	CG	66	117	159	CD	294	195	203	C	265	164	244
C	275	98	220	C	55	98	128	OE1	298	189	193	O	262	175	240
O	272	93	209	O	64	94	121	NE2	301	206	207	37 ILE			
9 SER				19 SER				N	261	207	230	N	265	153	237
N	285	93	228	N	43	96	126	C	261	207	230	CA	257	155	224
CA	290	80	224	CA	40	91	113	O	254	217	227	CB	265	151	211
CB	302	80	217	CB	25	91	108	29 TRP				CG2	256	154	199
OG	312	86	225	OG	16	93	119	N	278	218	230	CG1	278	158	208
C	287	69	235	C	48	97	102	CD1	288	156	219	C	244	147	224
O	295	60	236	O	50	91	91	O	244	134	226	O	244	134	226

TABLE 1 (continued)

10x 10y 10z				10x 10y 10z				10x 10y 10z				10x 10y 10z			
38	GLU			C	149	227	176	51	LYS			57	SER		
N	234	154	223	O	161	230	173	N	236	244	204	N	177	117	67
CA	220	148	223					CA	234	244	189	CA	183	106	61
CB	210	156	231	45	THR			CB	247	247	182	CB	197	110	55
CG	200	146	235	N	138	234	177	CG	258	240	188	OG	200	103	43
CD	191	153	243	CA	137	249	176	CD	271	243	181	C	185	95	72
OE1	183	162	237	CB	126	254	185	CE	263	236	187	O	190	98	83
OE2	189	151	256	CG2	121	244	195	N2	296	240	181				
C	215	146	208	OG1	131	265	193	C	227	232	183	58	GLU		
O	213	155	201	O	150	255	182	G	224	222	189	N	182	83	68
				O	154	251	193					CA	179	73	79
39	ARG							52	LEU			CB	174	58	73
N	214	133	206	46	VAL			N	225	233	169	CG	159	51	77
CA	208	128	193	N	157	263	175	CA	216	223	162	CD	145	60	77
CB	215	114	189	CA	170	268	179	CB	204	221	167	OE1	146	72	74
CG	230	114	191	CB	175	278	168	CG	193	229	159	OE2	133	55	80
CD	237	108	179	CG1	189	283	172	CD1	182	220	153	C	189	73	90
NE	251	106	182	CG2	177	271	154	CD2	185	239	167	O	186	77	101
C2	256	108	194	C	169	274	193	C	221	222	147				
NEH2	269	106	197	O	159	278	198	O	219	231	139	59	VAL		
NEH1	249	113	204					53	SER			N	202	69	89
C	193	124	196	47	LYS			N	227	210	144	CA	211	70	100
O	190	119	207	N	182	273	199	CA	228	203	131	CB	209	58	108
				CA	184	275	213	CB	243	198	130	CG1	208	61	123
40	GLY			CB	177	264	220	OG	246	198	116	CG2	198	49	105
N	186	128	186	CG	172	269	232	C	218	192	130	C	222	80	96
CA	171	126	186	CD	166	258	239	G	215	185	139	O	232	75	90
C	165	128	173	CE	161	263	252								
G	172	129	163	N2	156	252	260	54	CSS			60	CSS		
				C	195	281	216	N	210	195	117	N	220	92	100
41	CSS			O	206	277	211	CA	198	185	114	CA	228	103	95
N	152	128	172					CB	184	192	115	CB	219	113	86
CA	143	130	160	48	PRO			SG	179	199	132	SG	201	118	91
CB	130	123	158	N	196	293	226	O	199	178	101	C	233	111	107
SG	127	105	162	CD	184	298	232		202	183	91	O	237	123	105
C	138	144	157	CA	208	301	229	55	CSS			61	ASN		
C	131	150	166	CB	204	312	238	N	196	165	103	N	231	107	119
				CG	189	310	240	CA	197	155	93	CA	230	112	131
42	GLY			C	220	293	234	CB	208	147	92	CB	220	106	138
N	141	151	146	O	219	287	245	SG	205	134	103	CG	205	114	139
CA	134	164	143					C	186	145	88	OD1	196	111	132
C	143	176	144	49	GLY			O	178	138	96	ND2	204	125	146
O	154	176	147	N	229	293	225					C	243	109	140
				CA	243	287	227	56	GLU			O	242	100	149
43	CSS			C	243	272	225	N	182	145	76				
N	137	188	141	O	253	267	220	CA	170	140	69	62	ASN		
CA	145	200	142					CB	164	150	59	N	254	115	136
CB	149	206	128	50	ILE			CG	160	162	65	CA	266	114	144
SG	164	215	128	N	232	266	229	CD	154	172	55	CB	275	104	137
C	141	208	154	CA	230	251	227	OE1	149	183	60	CG	268	95	126
O	132	217	152	CB	218	246	235	OE2	153	170	43	OD1	269	96	114
				CG2	214	256	245	C	173	128	59	ND2	260	85	131
44	PRO			CG1	206	243	226	O	174	131	46	C	273	128	145
N	147	205	165	CD1	207	231	217					O	266	138	144
CD	155	193	166	C	227	249	212								
CA	145	212	177	O	215	251	208								
CB	153	204	187												
CG	159	192	180												

Molecular Conformation. Atomic coordinates for the 2.5 Å structure are given in Table 1. The Ramachandran angle plot (4) and a stereo drawing of the structure based on the coordinates are shown in Figs. 1 and 2 respectively. These results were obtained using the GRIP-75 Molecular Graphics System build by the University of North Carolina, Department of Computer Science. Fitting of the molecular model to the electron density map and idealization procedures were employed in an iterative mode to be described elsewhere. In the Ramachandran plot (Fig. 1), apart from the less restrained Gly angles, most residues fall within the outer limit boundary shown and all others are within at least 22° of this region. The three principal molecular lobes may

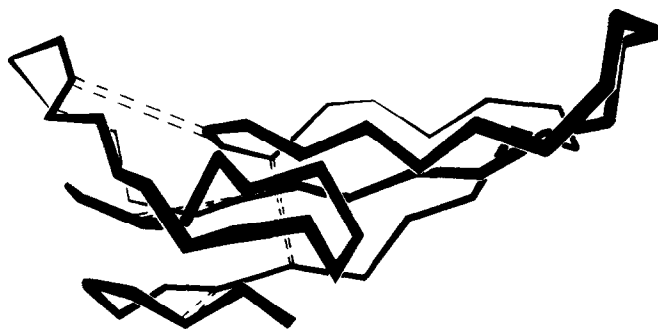


Fig. 3. Erabutoxin b. Freehand sketch of alpha-carbon backbone model. The double broken lines indicate disulfide bridges.

be seen to emerge in Fig. 2 from the central invariant disulfide core. These curved lobes, bounded by Cys3 and Cys17, Cys24 and Cys41 (K*45), and by Cys43 and Cys54 (K49, K61) are formed by the five BADCE strands of the dominant anti-parallel β structure (see inset) as well as by the outer rim segment 43-48 (K49-K55).

Fig. 3 shows the molecular shape in a view which emphasizes the curved shell-like concavity, and shows the protruding curled tongue of the long central DC lobe and the underlying "stand" formed by the C-terminal six-membered ring and 2 residue tail. In the short chain toxin series only the tapered neck region, Cys17-Cys24 (compare Fig. 4), is of variable length; all other molecular segments are invariant in length.

The erabutoxin b molecule is open, much of it only one peptide chain thick. Apart from a very few residues in the region around the disulfide core there are no buried residues or regions of backbone chain inaccessible to solvent. Nonetheless a striking degree of cross-linking maintains molecular stereochemistry. This includes not only hydrogen bonds between main-chain atoms in the β structure and elsewhere, but also numerous other main-chain and side-chain crosslinking interactions. Because of these interactions, the upper concave molecular surface (facing the viewer in Fig. 4) formed by the three

*The sequence enumerations used are that of erabutoxin b and, where different, that of Karlsson (K) which orders all sequence alignments to preserve maximum homology in both short and long chain series.

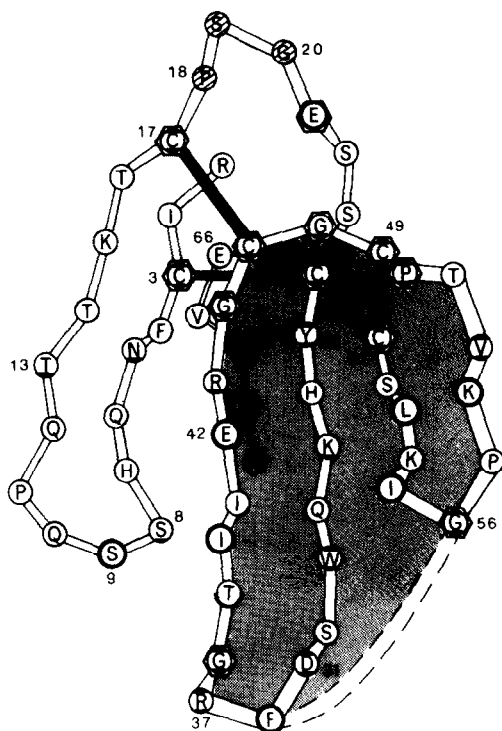


Fig. 4. View (partially schematic) of the erabutoxin b molecule looking into the concavity. The reactive site is shown shaded. One-letter residue code and Karlsson sequence enumeration used.

- Invariant position in both long and short chain toxins; in short chain toxins only ○ .
- Conservatively substituted position in both long and short chain toxins; in short chain toxins only ○ .
- ⬡ Structural residue position.

lobes is inaccessible even to water molecules from below, an isolation particularly significant for the reactive site region (5,6) shown shaded in the projected view.

Polypeptide Backbone. The barreling curve of the β structure is complex; individual strands both turn and twist. Fig. 5 shows the molecular hydrogen bonding pattern; the three β bends in the β sheet are, respectively, A7-B10 type I (7), C31-D34 (K38) type III (i.e., 3_{10} helix) and rim segment 47(K53)-E50 (K57) type II. There is a "wide" β bulge (8) in the left-hand edge of the sheet which involves Pro11 and Gln12 in strand B and Gln6 in the opposite A strand. The general preference for Pro in the first position has been

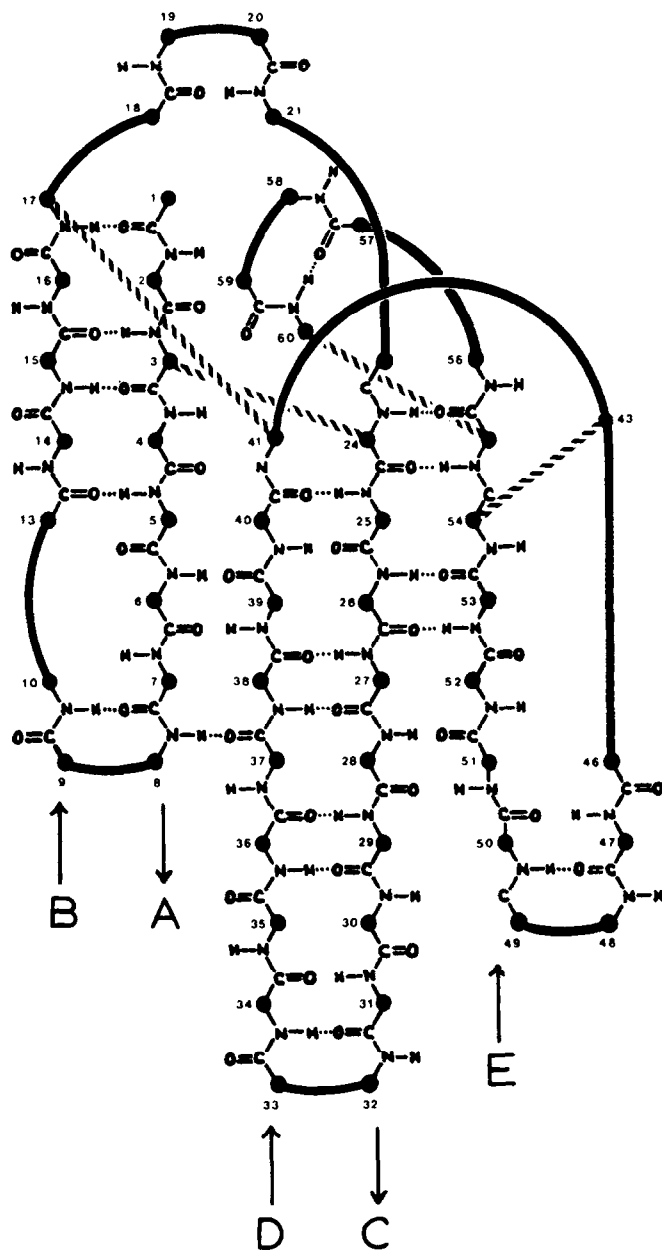


Fig. 5. Hydrogen bonding pattern of erabutoxin b polypeptide chain. The C=O and H-N groups are joined by dotted lines when the O-N distance ≤ 3.7 Å. Disulfide bridges are shown with striped lines.

noted earlier (8). Strands B and C are joined through the tapered neck region by a "plain +3 connection" which includes a type I β bend 18-21; strands D and E are joined by a right-handed $2\frac{1}{2}$ cross-over connection (9). The E strand leads into a type I β bend, 57-60 (K64-K68) in the C-terminal closed ring.

The dihedral angles about -S-S of the four disulfide bridges which maintain the β structure, all lie close to 90° . Three linkages are right-handed; that joining Cys17 and Cys41 (K45) is left-handed. The disulfide bridge 43-54 (K49-K61) is *Gauche.Gauche.Trans*; the other three are *Gauche.Gauche.Gauche*. These findings agree with the conclusion of the laser Raman solution studies of erabutoxin b (10).

Homologous Side Chain Distribution. The overall sequence homologies of invariant and conservatively substituted positions are further extended in the short series toxins by 6 more invariants and 2 more conservative substitutions. In a broad sense the extent and distribution of all common invariant or type-conserved residues define the principal features of the erabutoxin b structure as short series prototype and long series model. Most of these residues are found within the reactive site (1,2,5,6) region 32—49 ...32 (K33-K56...K33) formed by the two right-hand lobes of the molecule and shown shaded in Fig. 4. In the AB loop of the short-series toxins, the limited number of common residues may imply minor conformational variations; in the tapered neck region the one or two residue deletions found in positions 18,19, or 20 demand conformational variations.

There are residues which evidently play a key structural role; they lie within or are bound to the reactive site and are shown enclosed in open hexagons in Fig. 4. These include four invariant glycines, two in the β bends and two (one a short series invariant) at positions of close approach of two chains. Pro44 (K50) stabilizes the rim segment turn. Two further residues, the type-conserved position Glu21 (Glu/Asp) and the invariant Asn61 (K69) form hydrogen bonds to atoms in neighboring segments of the main chain.

A second group of residues has been designated "functional" because of probable involvement in receptor binding (5,6). With one exception, all point up along $C\alpha$ - $C\beta$ into the sweeping hollow of the reactive site. The exception Val46 (K52) forms part of the banked wall of this concavity. The functional residues have been sorted into reactive groupings (5,6). Toxicity

is less than normal when some functional residues are chemically modified or are found replaced in certain naturally occurring deviant toxins. Other hydrophobic residues have been designated "functional" on the basis of indirect evidence. We will consider the structure-function relationships and the effect of our recent findings on the details of the earlier model elsewhere.

The designations "structural" and "functional" are in some instances too restrictive; the two roles are not necessarily mutually exclusive. In a reactive grouping, one or more "functional" residues may position the residue which binds directly to receptor. Equally, some "structural" residues may be required for proper local functional expression, as for example, Gly34 (K38) in the acetylcholine mimetic association and binding of Asp31 and Arg33 (K37) proposed earlier (2,5,6,11).

It is a remarkable feature of the neurotoxin homology that functional residue distribution defines, and β structure stereochemistry demands a sequence distribution which involves an interrupted and frequently alternating series of invariant and/or conservative substitutions rather than a continuous segment of identical or near-identical sequence.

The atomic coordinates on which these conclusions are based may be expected to shift somewhat after refinement. Model studies have shown that differences between the long and short series toxins do not need to affect significantly the principal structural features we have described here (5,6).

ACKNOWLEDGEMENTS

This work was supported by grant NS-07747 from the National Institutes of Health. We happily acknowledge the use of GRIP-75 developed by E. G. Britton, F. P. Brooks, Jr., J. Hermans, J. S. Lipscomb, J. E. McQueen, M. E. Pique, and W. V. Wright.

REFERENCES

1. Low, B. W., Preston, H. S., Sato, A., Rosen, L. S., Searl, J. E., Rudko, A. D., and Richardson, J. S. (1976) Proc.Nat. Acad.Sci. U.S. 73, 2991-2994.
2. Low, B. W., Preston, H. S., Sato, A., Rosen, L. S., Searl, J. E., Rudko, A. D., and Richardson, J. S. (1979) Proceedings of the Fifth International Symposium Society of Toxinology, Costa Rica (1976), in Toxins - Animal,

- Plant and Microbial (P. Rosenberg, ed.) p. 312, Pergamon Press, N.Y.
3. Yang, C. C. (1967) *Biochim. Biophys. Acta* 133, 346-355.
 4. Ramakrishnan, C., and Ramachandran, G. N. (1965) *Biophys. J.* 5, 909-933.
 5. Low, B. W. (1979) in Snake Venoms-Handbook of Experimental Pharmacology (Chen-Yuan Lee, ed.) Vol. 52, pp 213-257, Springer-Verlag, Berlin.
 6. Low, B. W. (in press 1979) Proceedings of the Third International Symposium on Cytopharmacology in Advances in Cytopharmacology (B. Ceccarelli, F. Clementi, eds.) Vol. 3, pp. 141-147, Raven Press, New York.
 7. Venkatachalam, C. M. (1968) *Biopolymers* 6, 1425-1436.
 8. Richardson, J. S., Getzoff, E. D., and Richardson, D. C. (1978) *Proc. Nat. Acad. Sci. U.S.* 75, 2574-2578.
 9. Richardson, J. S. (1976) *Proc. Nat. Acad. Sci. U.S.* 73, 2619-2623.
 10. Harada, I., Takamatsu, T., Shimanouchi, T., Miyazawa, T., and Tamiya, N. (1976) *J. Phys. Chem.* 80, 1153-1156.
 11. Sato, A. (1977) Ph.D. thesis, Tohoku University, Japan.